



# Investigation of Optimum pH and Temperature for *In-Vitro* Crystallization of Urinary Cystine

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## ABSTRACT

*Cystinuria contributes in formation of urinary stones. But, it has been reported that cystinuria is diagnosed when someone experiences with cystine stones. Therefore, early diagnosis of this condition is important. Thus, the objective of the study was to determine the optimum pH and temperature for crystallization of urine cystine in-vitro. Cystinuria solutions were prepared with the concentrations of 40, 60, 70, 75, 80, 90, 100 and 120 mg/dL. The pH of each solution was changed with the addition of acetic acid. Then solutions were exposed to temperature +4°C and 37°C, for 15, 30 and 45min. The sediments were observed microscopically for cystine crystals formation. Then acetone was added to cystinuria with the ratio of cystinuria:acetone, 8:1, 4:1, 2:1 and 1:1 and pH was altered with acetic acid and were subjected to +4 °C and 37 °C, for 15, 30 and 45 minutes and sediment was observed for cystine crystals under the microscope. Cystine crystallization had been occurred in the cystinuria of  $\geq 100$  mg/dL at pH 5 at 37 °C and +4 °C, 30min after the addition of acetic acid whereas with the addition of acetone at cystinuria of  $\geq 75$ mg/dL at pH 5 in both 37°C and at +4°C, 30min after the addition of acetic acid. The number of cystine crystals per High Power Field (HPF) was highest where cystinuria:acetone was 8:1. The optimum conditions for cystine crystallization is at pH 5, 37 °C and +4 °C, 30min after acidifying with acetic acid at the minimum concentration of 100 mg/dL of cystinuria. With the addition of acetone, at the ratio of cystinuria:acetone 8:1 with minimum concentration of cystinuria of 75 mg/dL.*

**KEYWORDS:** Cystine, Crystallization, Acetic acid, Acetone, Temperature, pH



## 1. INTRODUCTION

Cystinuria is an autosomal-recessive inherited disorder in reabsorption and transport of cystine and the dibasic amino acids, ornithine, arginine and lysine from the luminal membrane of the renal proximal convoluted tubule and the small intestine, which eventually characterized by hyper-excretion of cystine and dibasic amino acids into urine. Eventually cystinuria plays a vital role in formation of urinary stones, being responsible for 1-2% urinary stones in the adult whereas up to 10% in children. Though, urolithiasis caused by cystine is the only phenotypic manifestation of cystinuria, it often persists throughout the life-time of the affected individual. It is a lifelong condition and if not managed and treated properly, cystinuria can be tremendously painful and may direct to serious medical complications such as damage to the kidney or bladder, urinary tract infections, pyelonephritis, ureteral and ureter obstructions.

Several investigations have been carried out on mineralogical compositions of urinary stones revealed that cystine as one of the contributor in urinary stone formation [Keshavarzi, 2016; Takasaki, 1995; Sakandé, 2012; Biyani & Cartledge, 2006]. Approximately 1% of adult and 6% of paediatric stones is cystine. It has been reported that estimated prevalence of cystinuria globally is 1 per 7000 individuals [Biyani & Cartledge, 2006]. It has identified that two genes are responsible for cystinuria condition. The cystinuria type I is caused by the mutations in the *SLC3A1* gene, located on the chromosome 2p, whereas non-type I cystinuria is due to variants in *SLC7A9*

[Jessen & Knoll; 2012, Biyani&Cartledge, 2006].

In addition to that the factors such as urine osmolality, dehydration, some food intake are also contributed in this condition. The reduced solubility of cystine in urine could be directed to stone formation in affected individuals. This would be manifested in the second and third decades of life. The management strategies are done to decline the urinary cystine concentration below 300 mg/L which involves plentiful oral fluid intake, urinary alkalinisation and thiol medications [Biyani&Cartledge, 2006].

The risk has been reported on recurrent formation of stones from cystine in the kidney, ureter and bladder. However, intermittent stone formation requires repetitive urological interventions with possibly impairment of renal functions and thereby quality of the life. The formation of cystine stone is initiated from neonatal stage and children are more vulnerable group to have cystine stones [Tasian & Copelovitch, 2014].

It has been reported that cystinuria is usually diagnosed when someone experiences with an incident of cystine stones. The diagnosis is usually made by testing the stones to demonstrate that they are made out of cystine. Despite the fact that early diagnosis of cystinuria is extremely important, due to increased solubility of cystine in urine of cystine stone formers compared to normal individuals [Nakagawa, 2000] cystine crystals may absent in urine deposit even in stone formers owing to miss-diagnosis of cystinuria.

Therefore, it has become very important to identify cystinuria condition by improving and modifying microscopic screening of

cystine crystal as a diagnostic test in urine sediment even under lower urine cystine concentrations to facilitate the early diagnosis before making complications. Hence, the objective of the present study was to determine the optimum pH and temperature for crystallization of urinary cystine *in-vitro*.

## 2. MATERIALS AND METHODS

### Preparation of cystine standard solution

Concentration series of cystine standard solution was prepared starting from the minimum cystine concentration which gives a positive result for cyanide nitroprusside test. The cystine standard solution of 75 mg/dL was prepared by dissolving 75 mg of cystine in 100 mL of 100 mmol/L  $\text{Na}_2\text{CO}_3$  solution. Then different concentrations of cystine

standards were prepared as given in Table 1.  $\text{NaCO}_3$  solution with 100 mmol/L was prepared by dissolving 1.06 g of  $\text{NaCO}_3$  in 100 mL distilled water.

### Preparation of cystinuria solution

Cystine standard solution was mixed with calculated volume of freshly voided urine sample which was collected from a 6 years old female healthy subject. According to the physical, biochemical and microscopic examination the urine sample was identified as a normal urine sample. The pH value of urine sample was measured by pH meter prior to add cystine standard solution. Concentration gradient of cystinuria solutions was 40, 60, 70, 75, 80, 90, 100 and 120 mg/dL. Preparation of cystinuria solutions was done according to the Table 1.

**Table 1:** Preparation of concentration gradient of cystinuria solution

Concentration of cystine standard solution	Volume of cystine standard solution	Volume of freshly voided urine	Final volume of cystinuria	Concentration of cystinuria
(40x5)=200 mg/dL	20 mL	80 mL	100 mL	40 mg/dL
(60x5)=300 mg/dL	20 mL	80 mL	100 mL	60 mg/dL
(70x5)=350 mg/dL	20 mL	80 mL	100 mL	70 mg/dL
(75x5)=375 mg/dL	20 mL	80 mL	100 mL	75 mg/dL
(80x5)=400 mg/dL	20 mL	80 mL	100 mL	80 mg/dL
(90x5)=450 mg/dL	20 mL	80 mL	100 mL	90 mg/dL
(100x5)=500mg/dL	20 mL	80 mL	100 mL	100 mg/dL
(120x5)=600 mg/dL	20 mL	80 mL	100 mL	120 mg/dL

For the preparation of 100 mL cystinuria solution with 75 mg/dL concentration, 80 mL of freshly voided urine was mixed with 20 mL of cystine standard solution with 375 mg/dL concentration. Because there was a 1:5 dilution when mixed 80 mL urine with 20 mL cystine standard solution and concentration of cystine standard solution fall off with 5 times from its initial value. Therefore, to prepare cystinuria with 75 mg/dL concentration,

the initial concentration of cystine standard solution was adjusted to 375 mg/dL (75mgx5).

After preparing cystinuria concentration series, pH value of each was measured by a calibrated pH meter. Then, aliquot from each cystinuria solution was centrifuged at 2000 rpm for 5 minutes and the sediment was observed microscopically for formation of cystine crystals.

**Table 2:** pH and temperature changes done on cystinuria solution

Temperature		pH value				
37 °C	Just after the acidification	6	5.5	5	4	3
	After 15 min	6	5.5	5	4	3
	After 30 min	6	5.5	5	4	3
	After 45 min	6	5.5	5	4	3
+4 °C	Just after the acidification	6	5.5	5	4	3
	After 15 min	6	5.5	5	4	3
	After 30 min	6	5.5	5	4	3
	After 45 min	6	5.5	5	4	3

#### **Determination of effect of temperature on crystallization of cystine *in-vitro***

The temperature was altered by keeping the cystinuria solutions in refrigerator (+4 °C) and at ambient temperature (37 °C), for different time periods; ie., 15 minutes, 30 minutes and 45 minutes. After that, cystinuria solution from each concentration was treated with acetic acid.

#### **Determination of effect of pH on crystallization of cystine *in-vitro***

The pH of prepared cystinuria solutions were adjusted to 6, 5.5, 5, 4 and 3 by acidifying the sample with acetic acid. All the pH measurements were taken by a calibrated pH meter.

Table 2 explains that how each cystinuria solution was treated with different pH values and temperatures. Before and after treating with each condition mentioned above, cystinuria solutions were observed macroscopically for the formation of precipitate. Then the samples were centrifuged at 2000 rpm for 5 minutes and the sediment was observed microscopically for formation of cystine crystals. The pH and the temperature which initiate the formation of cystine hexagonal plates were taken as the optimum conditions for cystine crystallization.

**Determination of effect of acetone on crystallization of cystine *in-vitro***

Acetone was added to each cystinuria solution according to the Table 3. Then pH and temperature were altered according to the Table 2 and urine sediment obtained from each concentration was observed under the microscope for the cystine crystal formation.

**Table 3:** Addition of acetone into the cystinuria solutions

Volume of cystinuria solution	Volume of acetone
5 mL	5.00 mL
5 mL	2.50 mL
5 mL	1.25 mL
5 mL	0.625 mL

All the tests were done in duplicate.

**3. RESULTS**

The initial pH of the freshly voided urine sample which used for the preparation of cystinuria solution was slightly acidic (pH=5.65). After preparing cystinuria concentration series, initial pH of fresh

urine shifted in to alkaline pH as indicated in Table 4.

**Table 4:** Initial pH values of cystinuria solutions

Concentration of cystinuria solutions	Initial pH
40 mg/dL	7.52
60 mg/dL	7.54
70 mg/dL	7.63
75 mg/dL	8.5
80 mg/dL	8.07
90 mg/dL	8.03
100 mg/dL	7.51
120 mg/dL	7.40

**Effect of pH and temperature on crystallization of cystine *in-vitro***

According to the microscopic observation, cystine crystals were present in the concentrations of  $\geq 100$  mg/dL of cystinuria solutions. Cystine crystallization had been occurred at pH 5 in both, 37 °C and at +4 °C, 30 minutes after the addition of acetic acid (see Table 5).

**Table 5:** Optimum pH and temperature for crystallization of urinary cystine *in-vitro*

Temperature	pH values	pH values				
		6	5.5	5	4	3
37 °C	Just after acidified	-	-	-	-	-
	After 15 min	-	-	-	-	-
	After 30 min	-	-	√	-	-
+4 °C	Just after acidified	-	-	-	-	-
	After 15 min	-	-	-	-	-
	After 30 min	-	-	√	-	-

Concentration of cystinuria solution = 100 mg/dL

- Cystine crystals were not present

√ Cystine crystals were present

### **Effect of acetone on crystallization of cystine *in-vitro***

With the addition of acetone, cystine crystallization had been occurred in the cystinuria solutions of  $\geq 75$  mg/dL at pH 5 in both 37 °C and at +4 °C, 30 minutes after the addition of acetic acid. Number of cystine crystals per HPF was highest in the solution where the ratio of cystinuria:acetone was 8:1. There was lesser number of cystine crystals where cystinuria: acetone was 2:1 and 4:1 with the deposition of amorphous phosphate over the cystine crystals. But there was no crystal formation in the solution where cystinuria:acetone was 1:1.

## **4. DISCUSSION**

Cystinuria is a metabolic disorder which is an autosomal-recessive defect in reabsorption, transport of cystine and the dibasic amino acids, ornithine and arginine, plays a vital role in forming urinary stones in adult and children [Jessen & Knoll, 2012; Fjellstedt *et al.*, 2003], but unfortunately can be identified only in few (19-26%) homozygous individuals. The risk has been reported due to recurrent formation of stones from cystine in the kidney, ureter and bladder though the condition is rare. The formation of cystine stone is initiated from neonatal stage and children are more vulnerable group to have cystine stones [Perera, 2016]. However, increased solubility of cystine has been reported from urine of

cystine stone formers than normal subject. Cystine crystals may absent in urine deposit even in stone formers owing to miss diagnosis of cystinuria.

According to the literature there are different methods in detecting heterozygous cystinuria such as cyanide-nitroprusside test, thin-layer amino acid chromatography, colorimetric estimations of cystine and ion-exchange amino acid chromatography. According to the results the highest sensitivity is given by thin-layer chromatography. The same study highlighted that the frequency of heterozygotes calculated in other studies, may be under-estimated which based on screening by the cyanide nitroprusside test. As a screening test the colorimetric estimations provides low sensitivity [Giuglian, 1987].

As the colour development of nitroprusside test from cystine is sometimes negative for stone formers, it has been suggested that nitroprusside procedure is not suitable for the screening and quantitative measurement of cystinuria [Wu, 1992]. So, it justified the need to develop a microscopic screening method for cystine. In the present study, the urine sample was collected from a healthy female of 6 years old in order to prepare cystinuria solutions. According to the physical, biochemical and microscopic examination this urine sample was identified as a normal urine sample. The urine sample was collected from a single person in order to minimize the other variables that could affect the study. Diminishing volume of acetone was added to lower the ionic strength of surrounding

by increasing the electro static forces, after previously just acidifying with acetic acid in order to lower the pH up to optimum PH=5. As acetone gives a precipitate, consisting of cystine crystals mixed with phosphates and oxalates it disturbed the clear visualization of hexagonal crystals due to deposition of phosphate over cystine which should be diminished by prior precipitation of phosphates with ammonia solution followed by calcium chloride solution before adding acetic acid.

## 5. CONCLUSION

The optimum conditions for cystine crystallization is; pH 5 at 37°C and +4°C, 30 minutes after acidifying with acetic acid at the minimum concentration of 100 mg/dL cystinuria and cystinuria:acetone 8:1 with minimum concentration of 75 mg/dL.

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## REFERENCES

Biyani, C.S. and Cartledge, J.J. (2006). Cystinuria-diagnosis and management. *EAU-EBU update series*, 4(5), pp.175-183.

Fjellstedt, E., Harnevik, L., Jeppsson, J.O., Tiselius, H.G., Söderkvist, P. and Denneberg, T. (2003). Urinary excretion of total cystine and the dibasic amino acids arginine, lysine and ornithine in relation to genetic findings in patients with cystinuria

treated with sulfhydryl compounds. *Urological research*, 31(6), pp.417-425.

Giugliani, R., Ferrari, I. and Greene, L.J., (1987). An evaluation of four methods for the detection of heterozygous cystinuria. *Clinicachimicaacta*, 164(2), pp.227-233.

Jessen, J.P. and Knoll, T. (2012). Management of Cystinuria. In *Urolithiasis* (pp. 757-765). Springer, London.

Keshavarzi, B., YavarAshayeri, N., Moore, F., Irani, D., Asadi, S., Zarasvandi, A. and Salari, M. (2016). Mineralogical composition of urinary stones and their frequency in patients: relationship to gender and age. *Minerals*, 6(4), p.131.

Nakagawa, Y., Asplin, J.R., Goldfarb, D.S., Parks, J.H. and Coe, F.L. (2000). Clinical use of cystinesupersaturation measurements. *The Journal of urology*, 164(5), pp.1481-1485.

Perera, I. (2016). Renal stones in children: evaluation and medical management. *Sri Lanka Journal of Child Health*, 45(1).

Sakandé, J., Djiguemde, R., Nikiéma, A., Kabré, E. and Sawadogo, M. (2012). Survey of urinary crystals identified in residents of Ouagadougou, Burkina Faso: Implications for the diagnosis and management of renal dysfunctions. *Biochimie*, 24(3), pp.123-128.

Takasaki, E., Suzuki, T., Honda, M., Imai, T., Maeda, S. and Hosoya, Y.( 1995). Chemical compositions of 300 lower urinary tract calculi and associated disorders in the urinary tract. *Urologiainternationalis*, 54(2), pp.89-94.

Tasian, G.E. and Copelovitch, L.( 2014). Evaluation and medical management of kidney stones in children. *The Journal of urology*, 192(5), pp.1329-1336.

Wu, J.T., Wilson, L.W. and Christensen, S. (1992). Conversion of a qualitative screening test to a quantitative measurement of urinary cystine and homocystine. *Annals of Clinical & Laboratory Science*, 22(1), pp.18-29.